

# LABORATORY OF MICROBIOLOGY AND CELLULAR BIOLOGY

*Prospecting Hydrolases of  
microorganisms and finding their  
potential for bioenergy production*

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# Presentation outline

**Biomass**

**Biofuels**

**Enzymes from microorganisms**

**Perspectives**

# Biomass

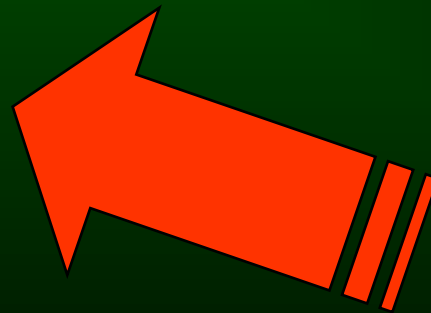
## What is it?

All renewable resources  
which come from organic  
matter and that can be  
used to produce energy.

Three domains  
of life and their  
wastes

2.000.000.000.000

tons



# Biofuels

**Solid, liquid or gaseous fuel**



wood, grass cuttings, domestic refuse, charcoal, dried manure, residues



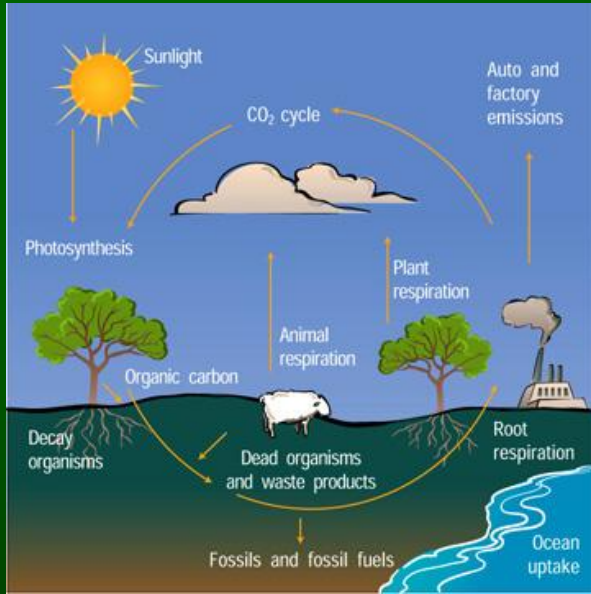
Vegetable oil, alcohol, biodiesel



Syngas, Biogas

## Advantages

# Biofuels

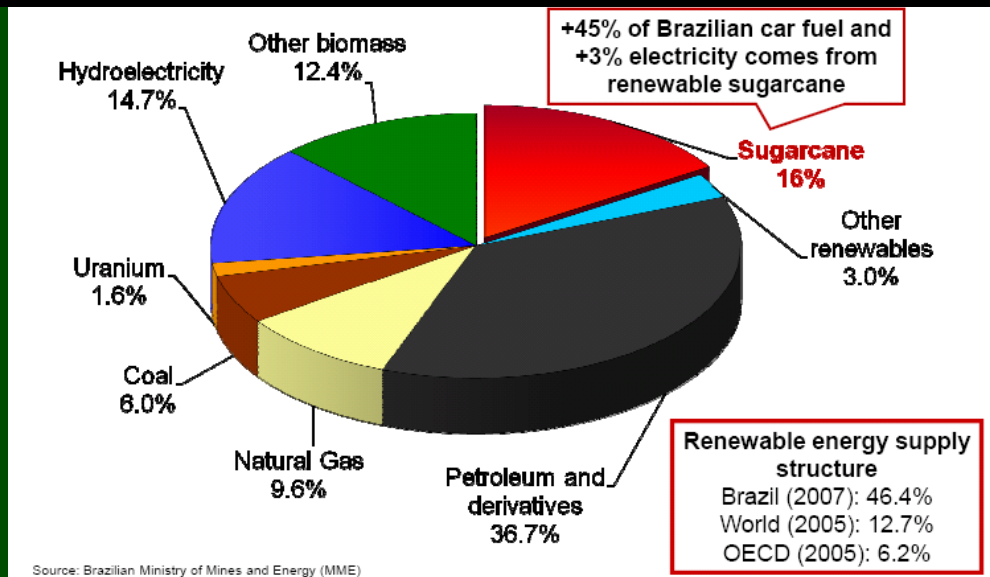


**GONE**

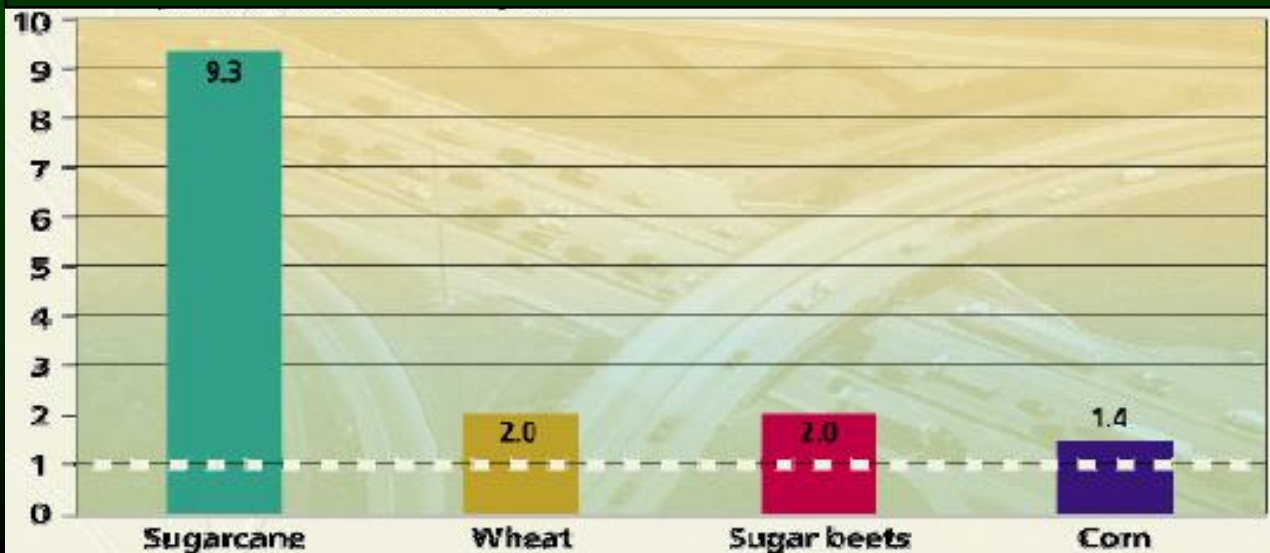




## Brazil Energy Matrix Input

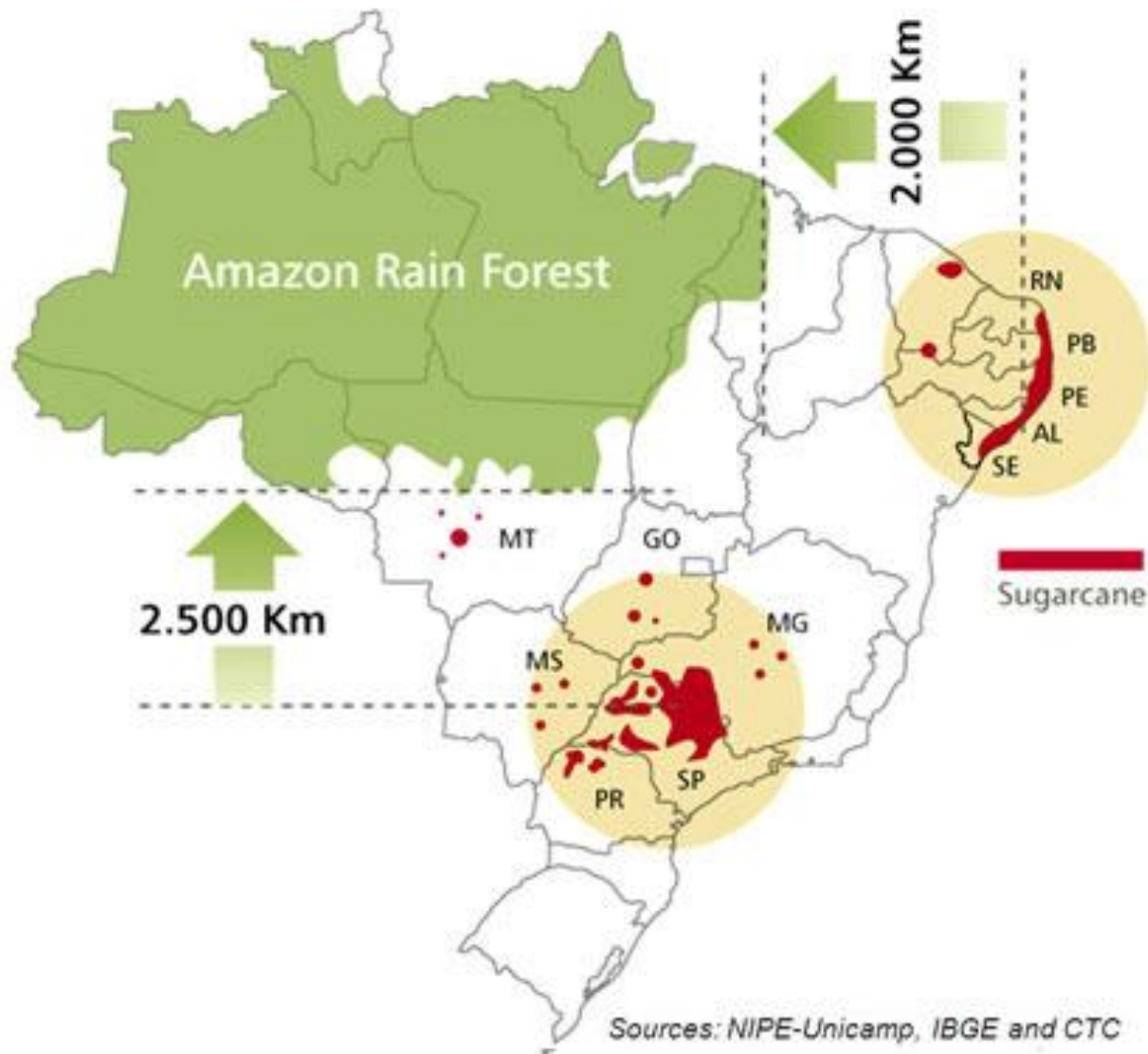


## SUPERIOR ENERGY BALANCE



Fonte:  
 World  
 Watch  
 Institute  
 (2005) and  
 Macedo et  
 al. (2008)

# Sugarcane producing regions in Brazil



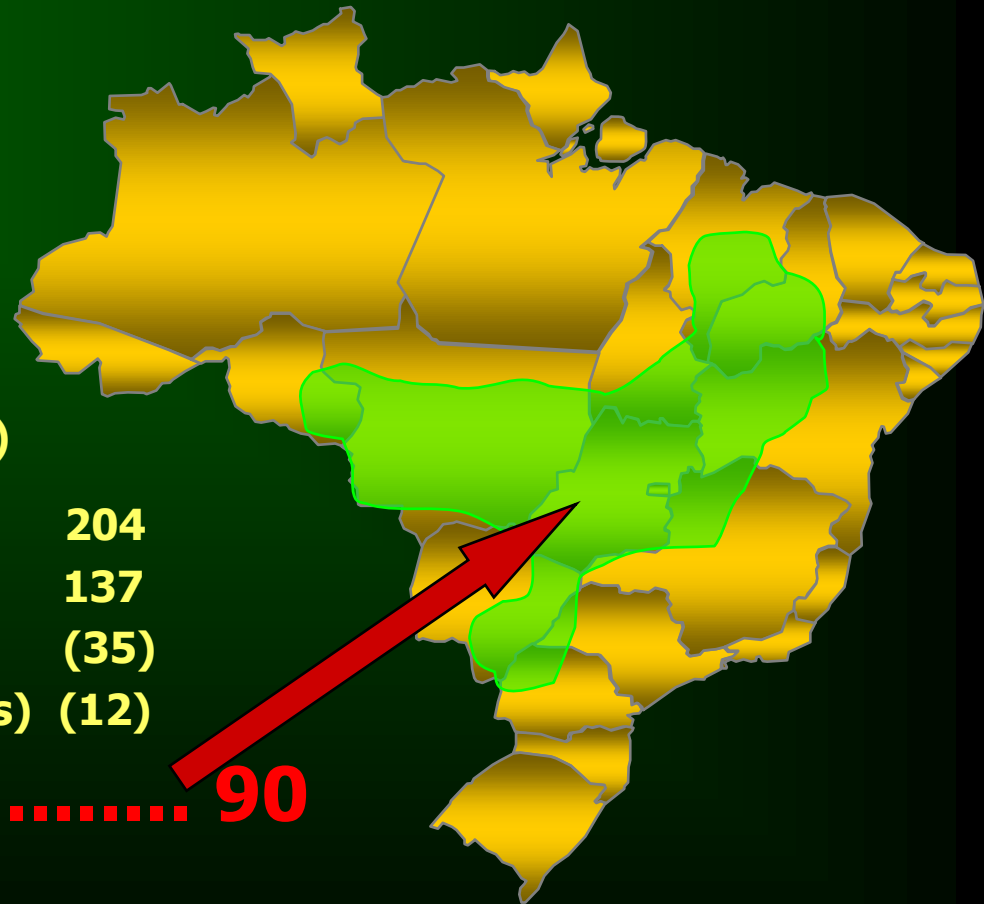
# BRAZIL: EXPANSION POTENTIAL

BRAZIL IS THE ONLY COUNTRY IN THE WORLD WITH GREAT CAPACITY TO EXPAND ITS AGRICULTURAL PRODUCTION (with sustentability)

Expansion area in the “Brazilian Cerrado” region

(million hectares)

Total Area .....	204
Area good for agriculture.....	137
Area in use for cattle raising...	(35)
Occupied area (forests & plantations)	(12)
<b>Available Area .....</b>	<b>90</b>



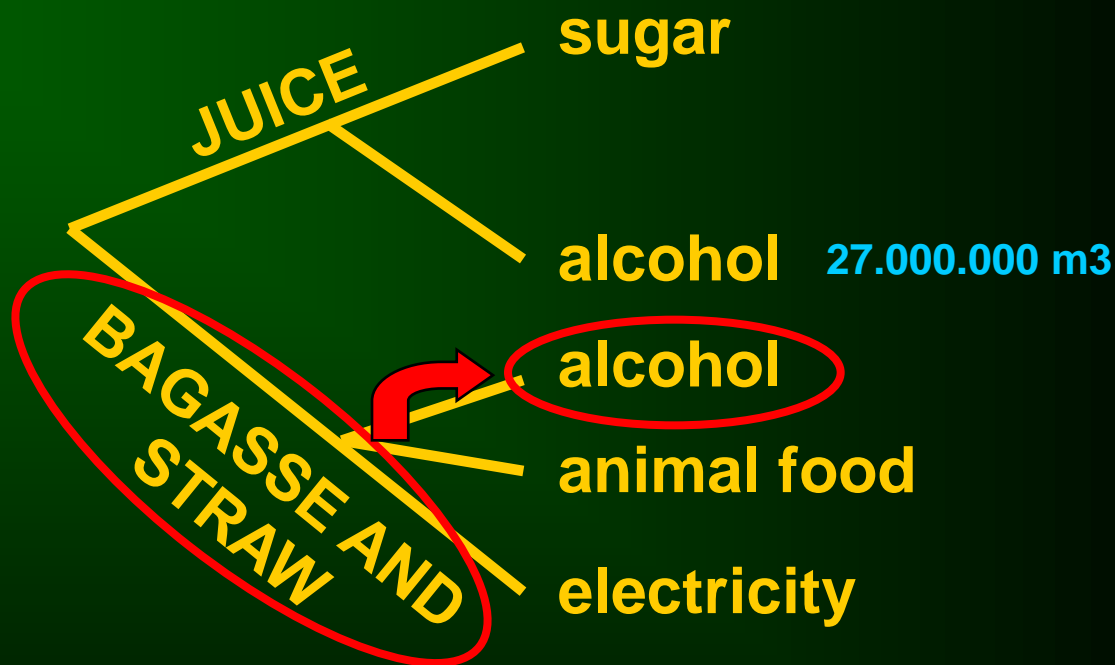


# Second generation ethanol



**Sugarcane**

**426 million tons**



# Ethanol

**It is biodegradable, made from renewable sources**

**Low toxicity to aquatic and soil forms of life**

**Virtually no sulfur and particulate emissions**

**Reduces emissions of CO and toxic substances**

**Blended with other fuels**

**Manufacture biodiesel (methanol)**

**Source of hydrogen (fuel cells)**

**Increasing Market of flexible fuel vehicles**

**Alternative fuel for motorcycles, boats, airplanes**

**Plastic production**

**Electricity**

# Cellulosic ethanol



600.000.000 tons



75.000.000 tons

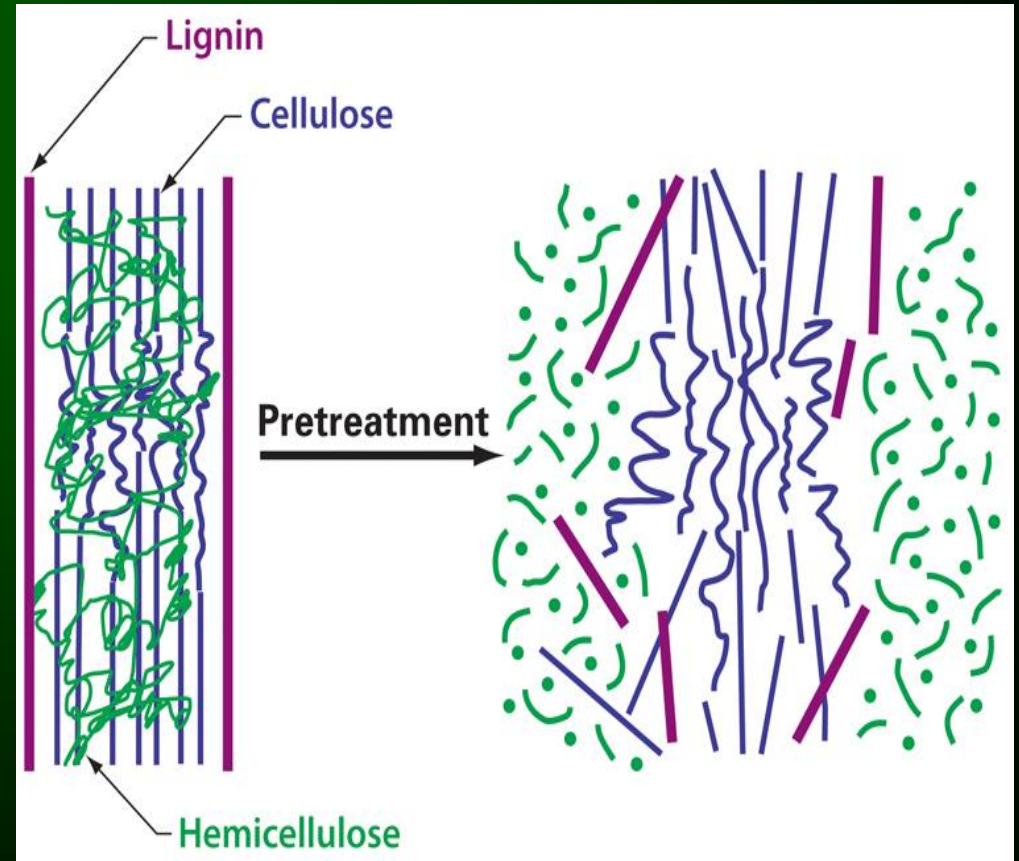


61.200.000 tons

10.000.000 tons

2.400.000.000 L

# Cell wall structure



# Enzymes:

<b>Amylases</b>	<i>Aspergillus phoenicis</i> , <i>A. niveus</i> ; <i>Rhizopus microsporus</i> var. <i>rhizopodiformis</i> ; <i>Paecilomyces variotii</i>
<b>Celulases</b>	<i>Aspergillus phoenicis</i> , <i>A. niveus</i> ; <i>A. japonicus</i> ; <i>A. ochraceus</i> ; <i>A. niger</i> ; <i>A. terricola</i> ; <i>Malbranchea puchella</i>
<b>Xylanases</b>	<i>Aspergillus phoenicis</i> , <i>A. niveus</i> ; <i>A. japonicus</i> ; <i>A. ochraceus</i> ; <i>A. niger</i> ; <i>A. terricola</i> ; <i>Malbranchea puchella</i>
<b>Xylosidases</b>	<i>Aspergillus phoenicis</i> , <i>A. niveus</i> ; <i>A. japonicus</i> ; <i>A. ochraceus</i> ; <i>A. niger</i> ; <i>A. terricola</i> ; <i>Malbranchea puchella</i>
<b>Ligninases</b>	<i>Aspergillus phoenicis</i> , <i>A. niveus</i> ; <i>A. japonicus</i> ; <i>A. ochraceus</i> ; <i>A. niger</i> ; <i>A. terricola</i> ; <i>Malbranchea puchella</i>
<b>Invertases</b>	<i>Aspergillus phoenicis</i> ; <i>A. caespitosus</i>
<b>Lipases</b>	<i>Trichoderma pseudokoningii</i> ; <i>A. caespitosus</i> ; <i>A. niger</i> ; <i>Cordyceps brongniartii</i> ; <i>Penicillium purpurogenum</i>
<b>Pectinases</b>	<i>Aspergillus terreus</i> ; <i>A. niveus</i> ; <i>Rhizopus microsporus</i> var. <i>rhizopodiformis</i> ; <i>Paecilomyces variotii</i>
<b>Acid and alkaline phosphatases</b>	<i>Aspergillus niger</i> ; <i>A. niveus</i> ; <i>A. ochraceus</i> .
<b>Phytases</b>	<i>Aspergillus niger</i> ; <i>A. niveus</i> ; <i>A. ochraceus</i> .



Syrup starch



Cheese preparation



Fruit juice



Pulp & paper



Animal food

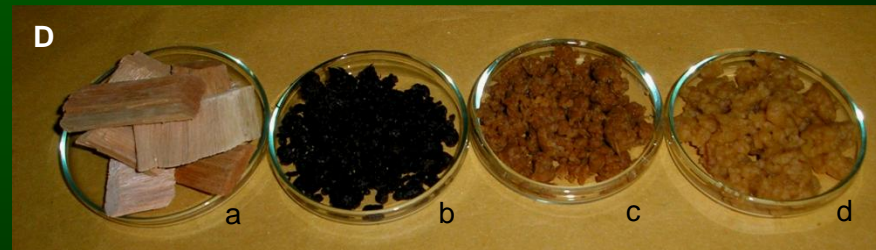


detergent



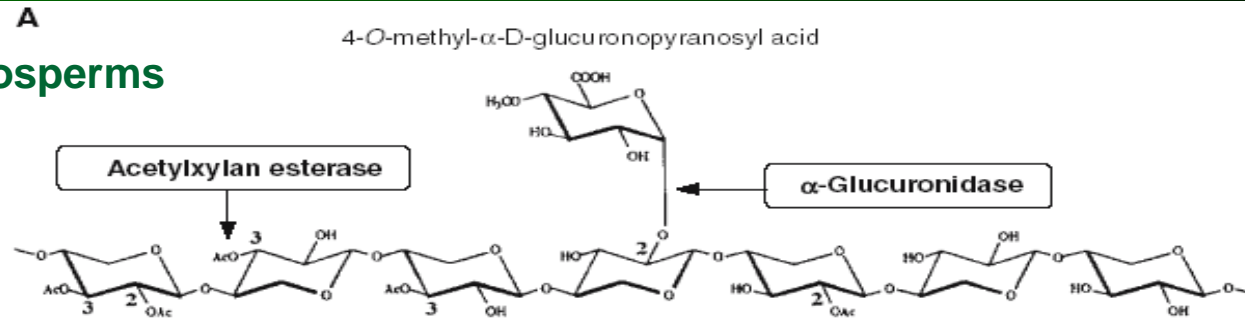
Ethanol

# BIODBLEACHING OF CELLULOSIC PULP

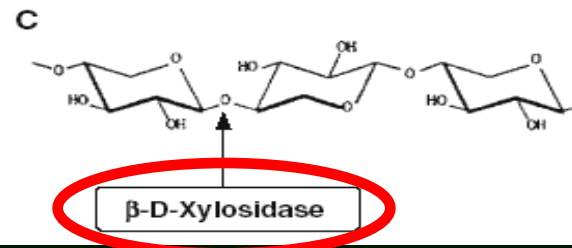
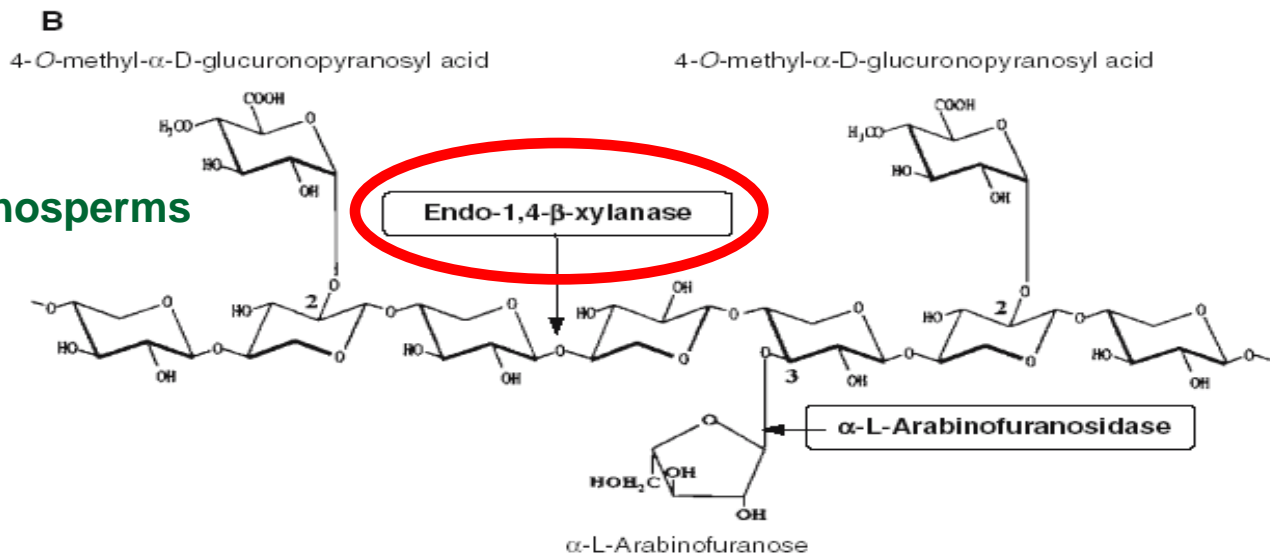


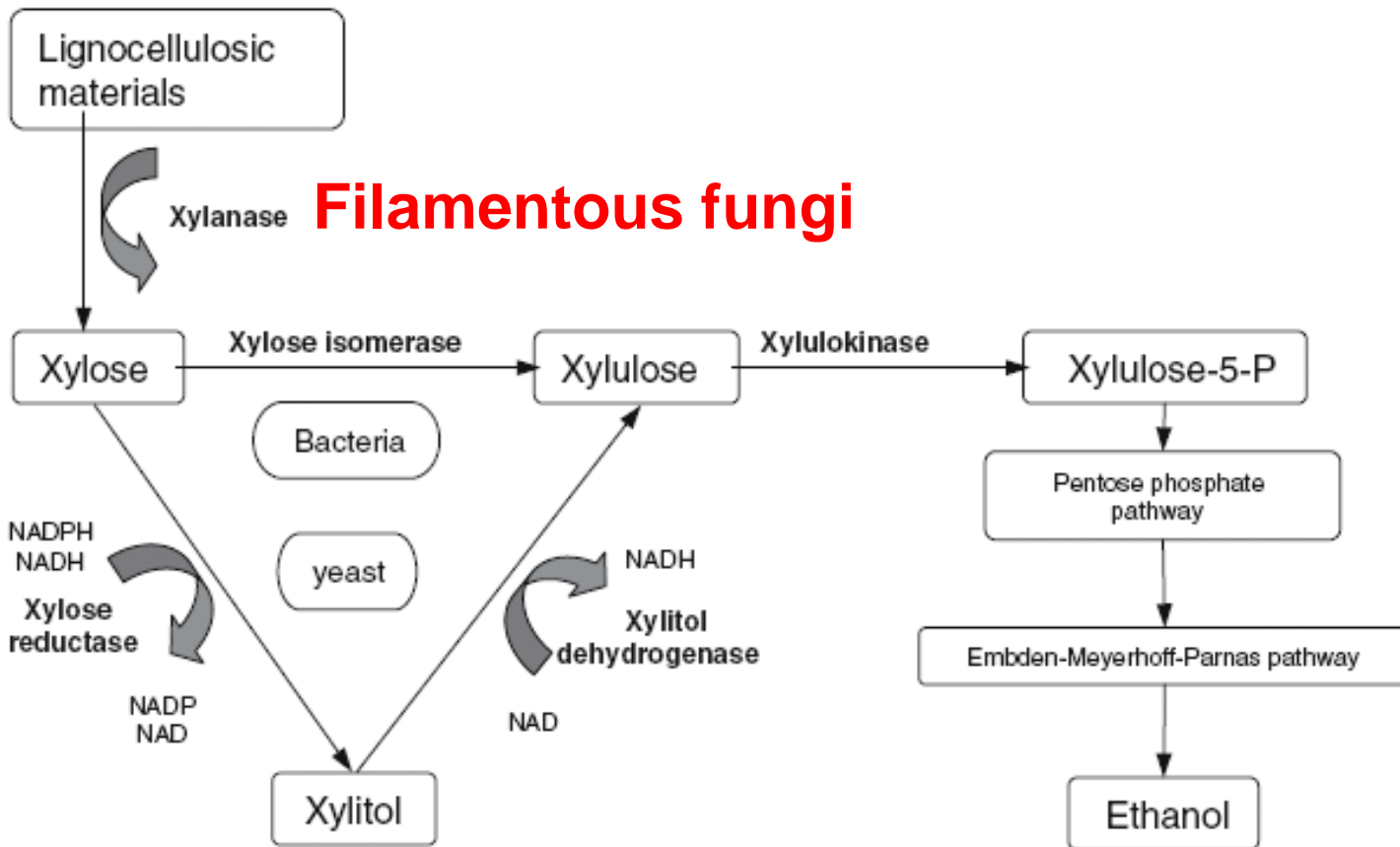
(f) NaOH/O<sub>3</sub>/ClO<sub>2</sub> (g) NaOH/O<sub>2</sub>/H<sub>2</sub>O<sub>2</sub> (h) NaOH/ClO<sub>2</sub>

## Angiosperms



## Gimnosperms





Polizeli, M.L.T.M. et al., Xylanases from fungi: properties and industrial applications. Review. *Appl. Microbiol. Biotechnol.*, 67, 577-591, 2005



# Collection of samples



beehive



Soil and leaves



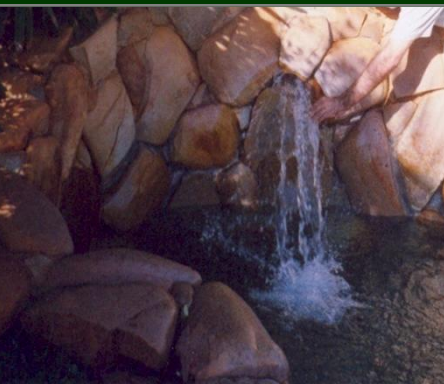
Sugar cane soil



Soil and humus of farm



edge of the river



Thermal water



Orange trees soil



Eucalipto forest



Maize plantation

# Fungus samples



# Fungus isolates



# Fungi storing



# Fungi characterization

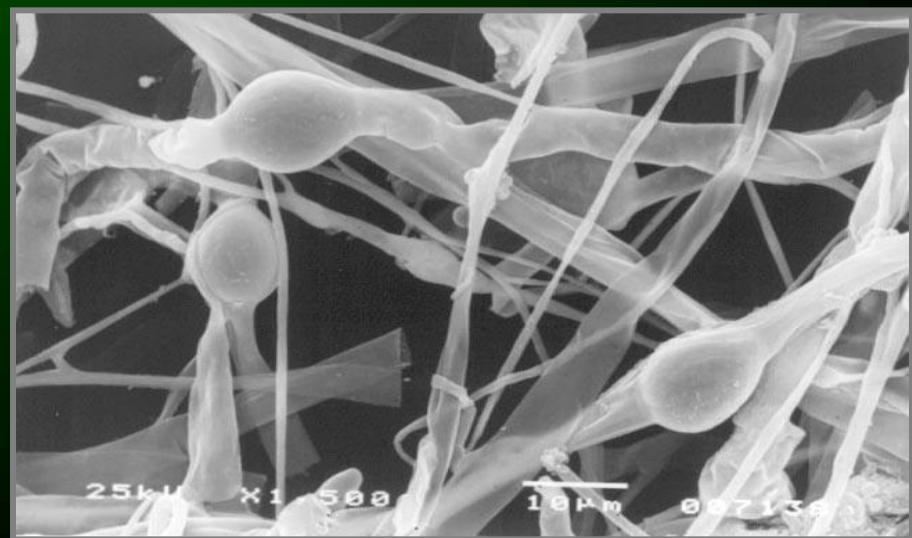
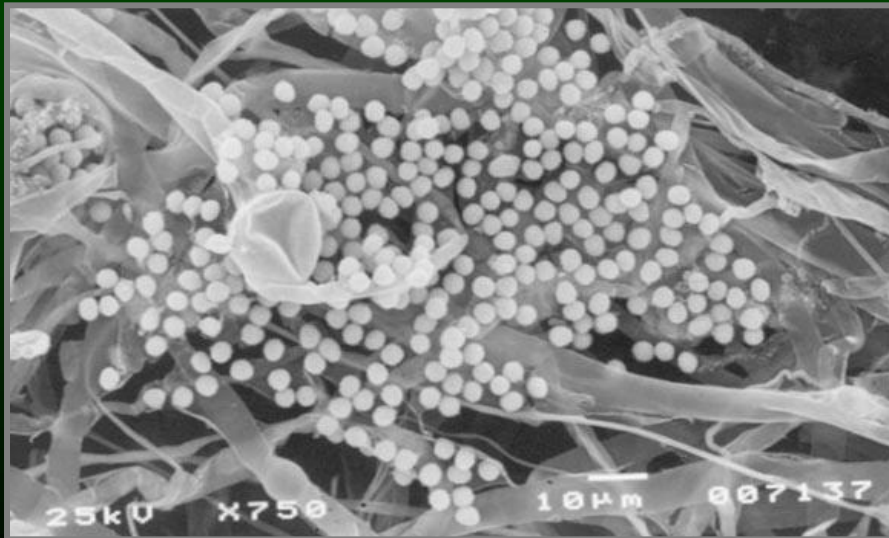
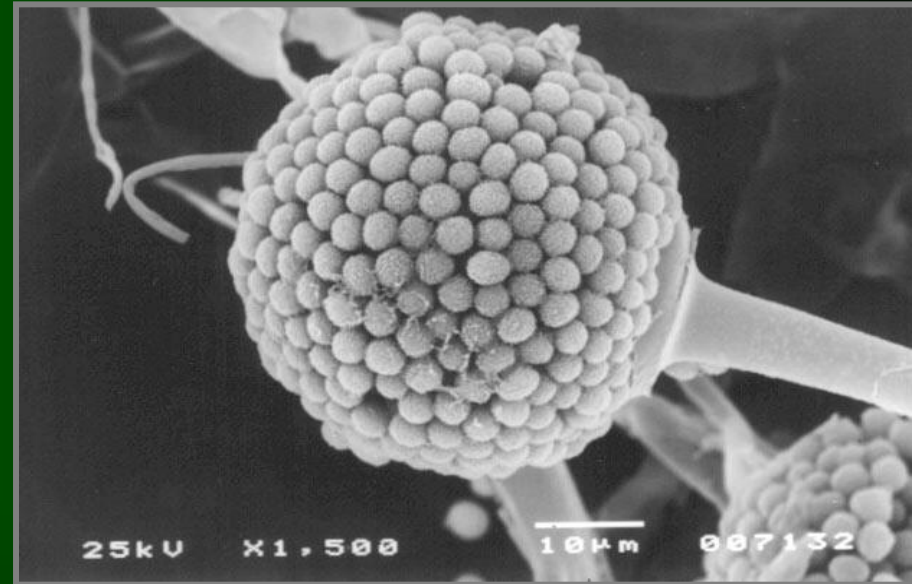
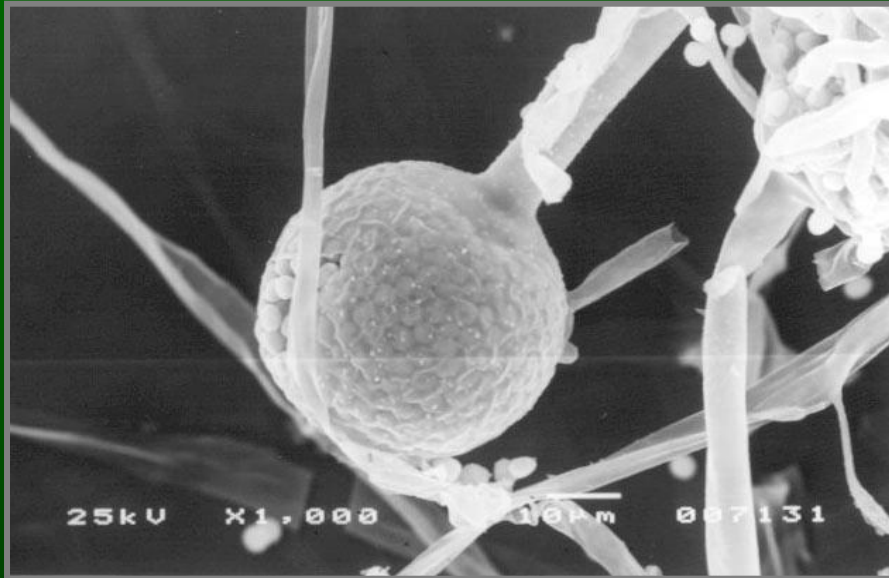


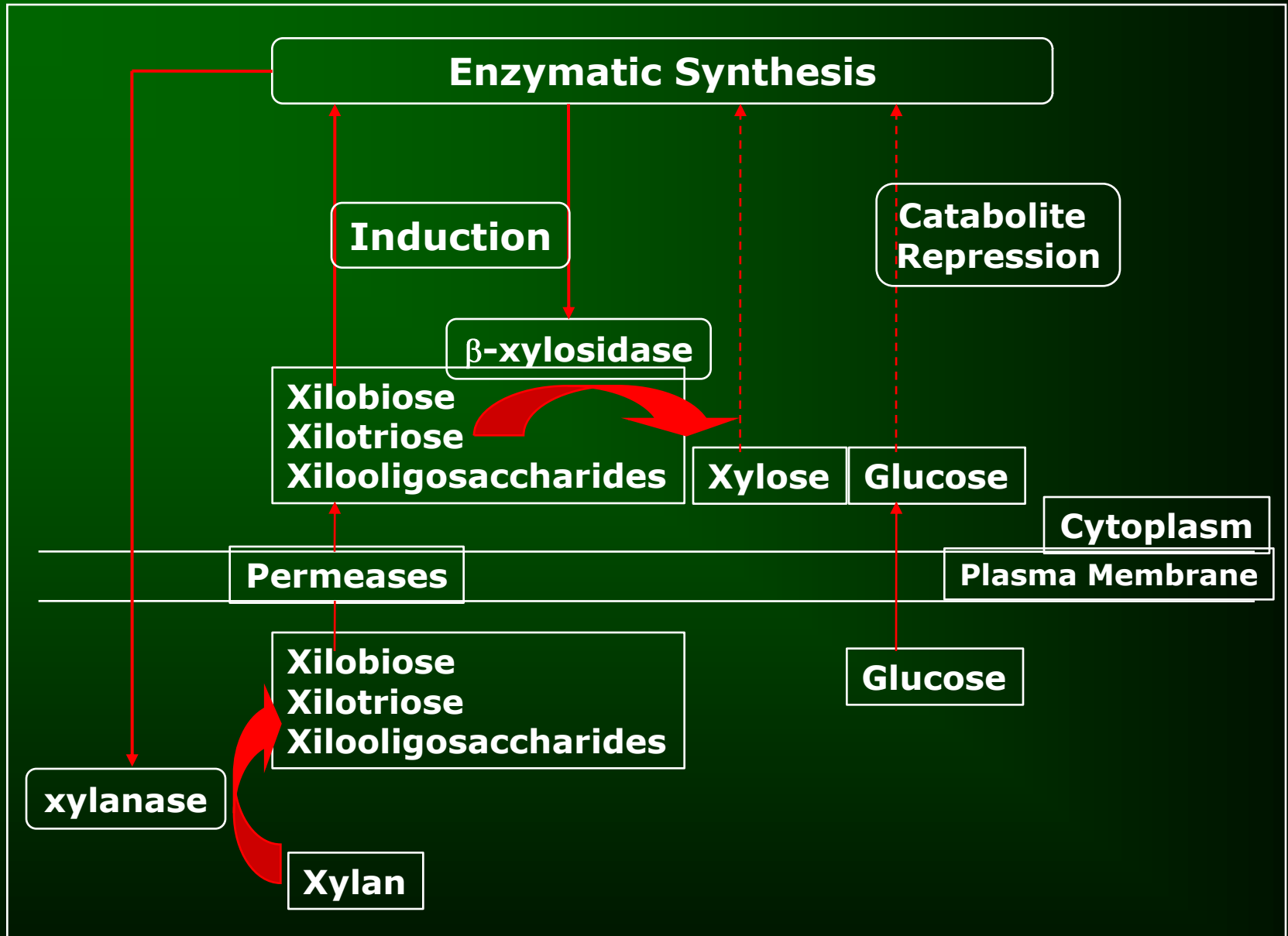
Brazilian Journal of Microbiology (2006) 37:474-480  
ISSN 1517-8382

## SCREENING OF FILAMENTOUS FUNGI FOR PRODUCTION OF ENZYMES OF BIOTECHNOLOGICAL INTEREST

Luis Henrique S. Guimarães<sup>1</sup>; Simone C. Peixoto-Nogueira<sup>2</sup>; Michele Michelin<sup>1</sup>; Ana Carolina S. Rizzatti<sup>1</sup>; Valéria C. Sandrim<sup>3</sup>; Fabiana F. Zanoelo<sup>1</sup>; Ana Carla M.M. Aquino<sup>1</sup>; Altino B. Junior<sup>2</sup>; Maria de Lourdes T.M. Polizeli<sup>1\*</sup>

# Fungi characterization





# Effect of carbon sources on $\beta$ -xylosidase and xylanase production

Carbon source	Concentration (%)	Protein (mg total)	Specific activity <sub>1</sub> U (mg protein)
None		1.5	8.6
Sugars			
Avicel	1	1.7	nd <sup>b</sup>
Cellobiose	2	1.8	95.4
Fructose	2	3.6	45.7
Glucose	2	4.7	34.5
Maltose	2	4.4	43.4
Raffinose	2	1.3	nd
Starch	1	1.6	82.9
Sucrose	2	1.9	nd
Xylan	1	2.3	278.3
Xylose	1	1.8	146.7
Industrial residues			
Bagasse sugar cane	1	2.9	219.9
Cassava flour	1	3.3	nd
Maize pith	1	4.0	112.3
Oatmeal	1	3.1	nd
Rice peal	1	2.7	nd
Wheat raw	1	5.2	35.0

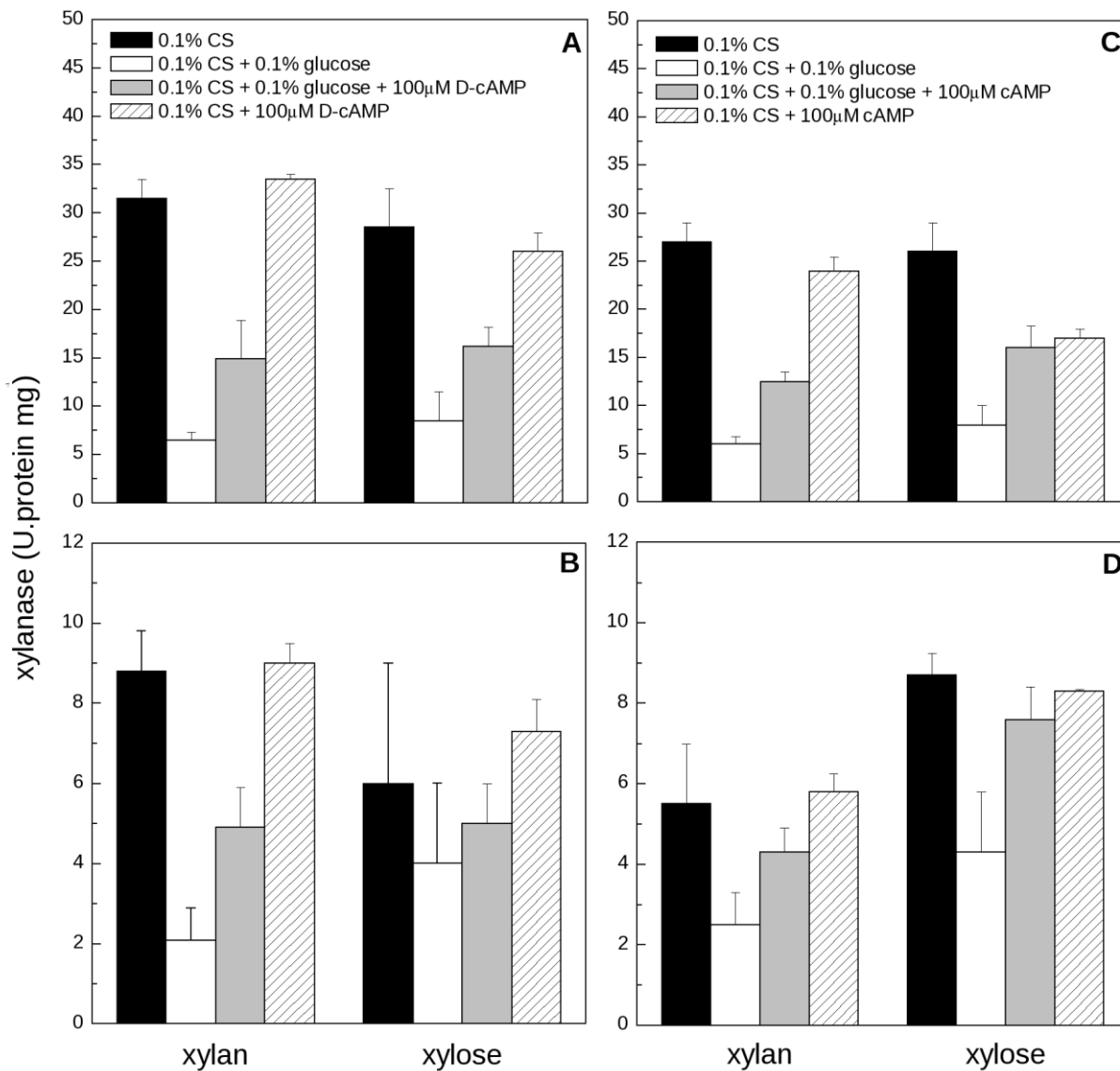
**Polizeli, M.L.T.M. et al., *J. Ind. Microb. & Biotechnol.* 26, 156-160, 2001.**

Carbon source (1%)	Extracellular xylanase (U mg protein <sup>-1</sup> )
Control <sup>a</sup>	1.15
Arabinose	8.90
Fructose	3.31
Galactose	1.70
Glucose	2.37
Mannose	2.50
Ribose	2.22
Xylose	152.00
Xylose + glucose	40.00
Cellobiose	1.52
Raffinose	1.42
Lactose	0
Maltose	5.90
Sucrose	4.14
Starch	13.10
Xylan (birchwood)	172.40
Xylan (oat spelt)	171.40
Xylan (beechwood)	173.00
Xylan + glucose	90.00
Xylitol	3.14
$\beta$ -Methylxyloside	152.30
$\beta$ -Methylxyloside + glucose	25.00

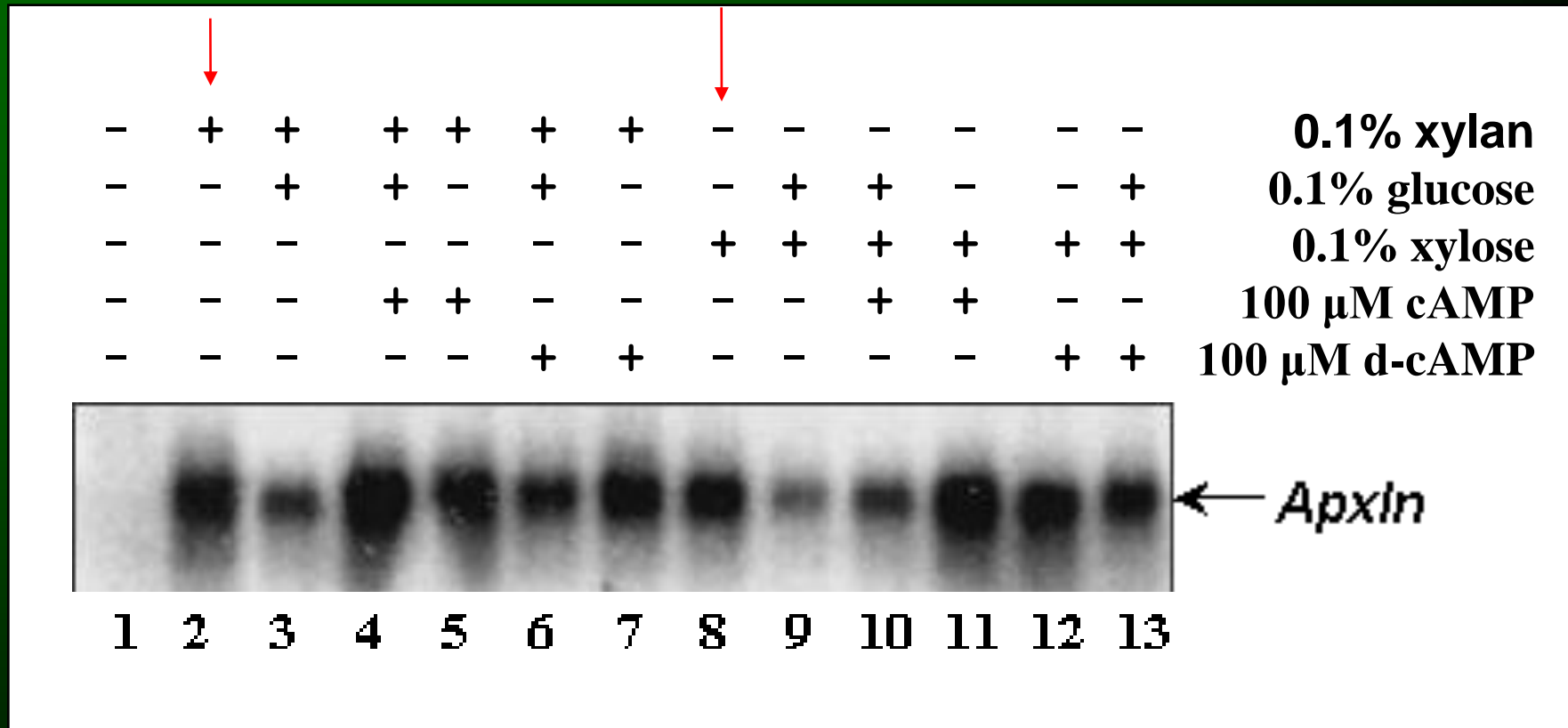
**Rizzatti, A. C. et al., *J. Ind. Microbiol. Biotechnol.* 35 (4), 237- 244,.2008.**



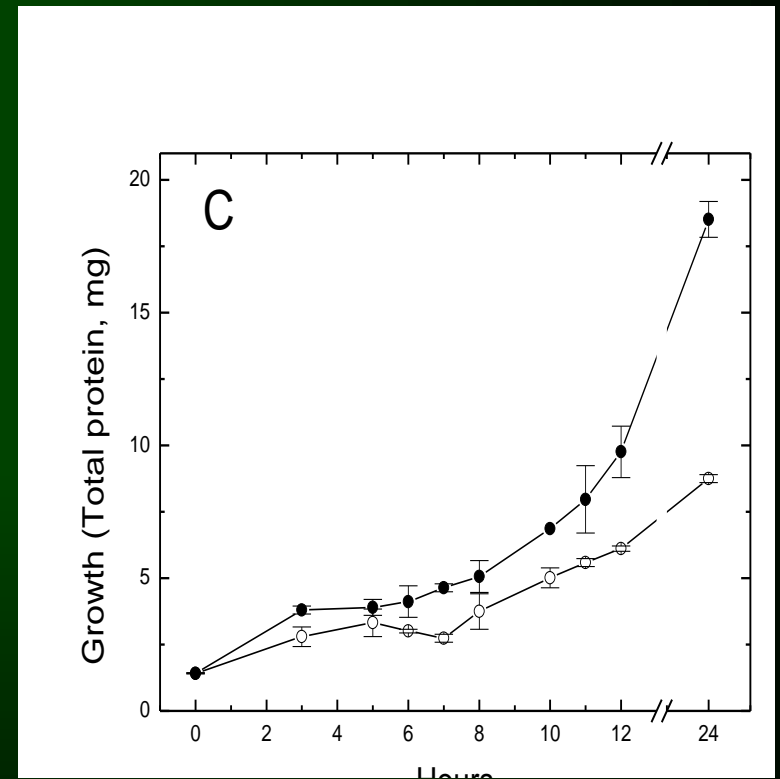
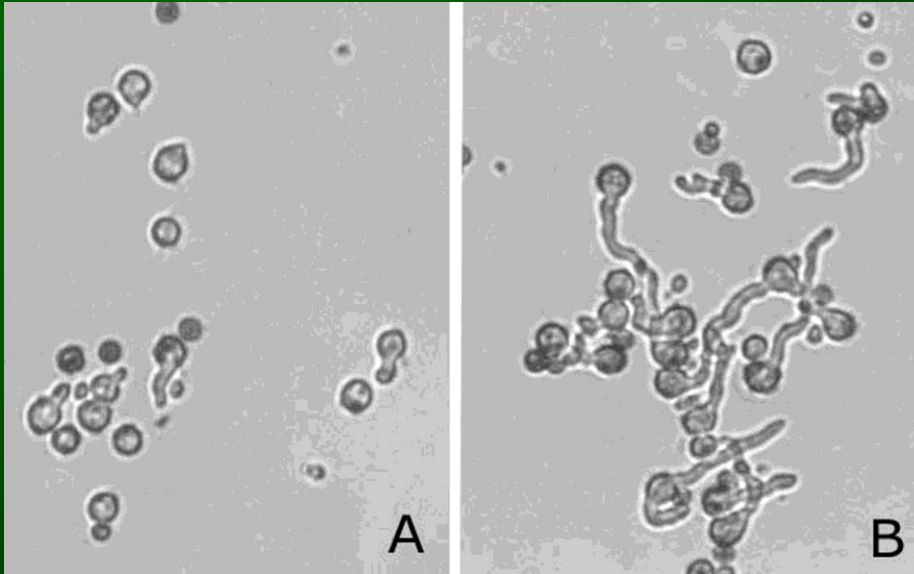
# Effect of glucose, dibutyryl-cAMP or cAMP on xylanase production



# Northern blot hybridization of the *A. phoenicis* gene *ApXLN*

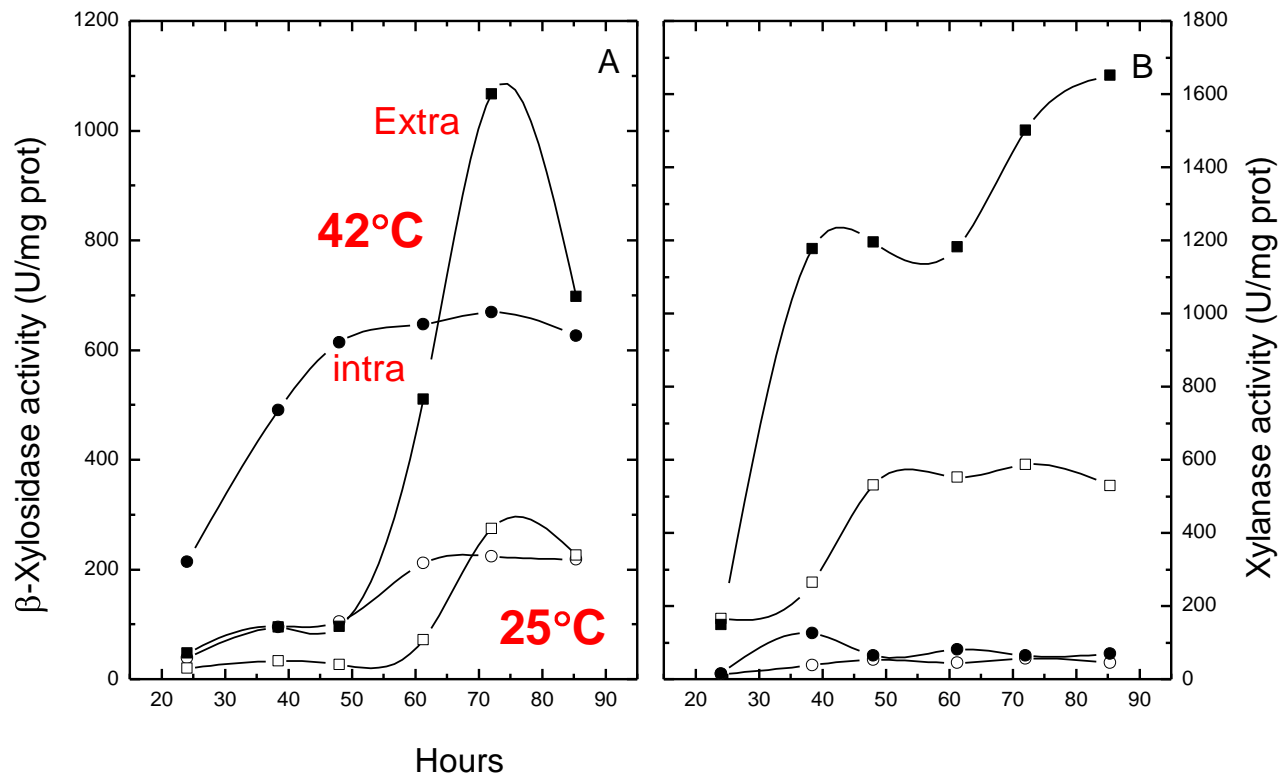


# Spore germination of *A. phoenicis* at 25 C (A) and 42 C (B) for 8 hs



(C) Growth at 25 C (O) and 42 C (●).

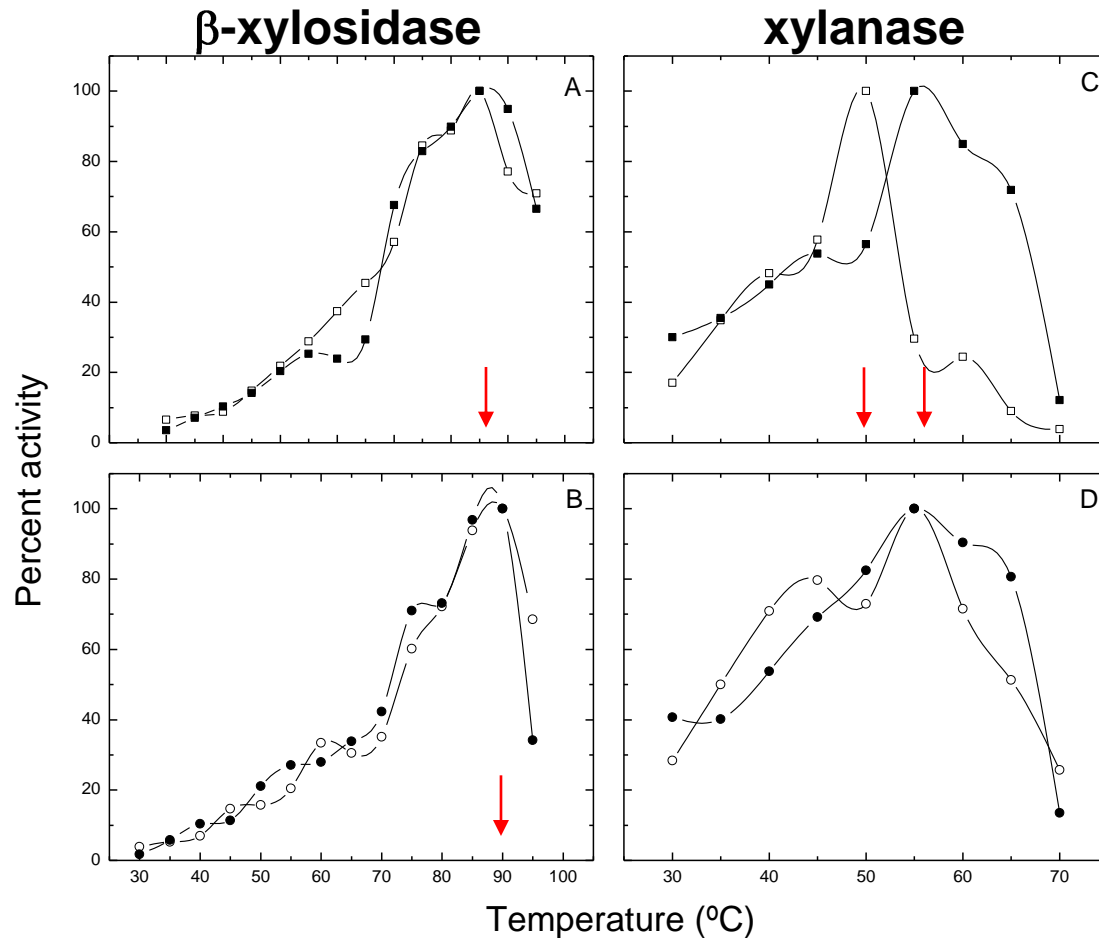
# Time-course of $\beta$ -xylosidase (A) and xylanase (B) production



# Effect of temperature on activities

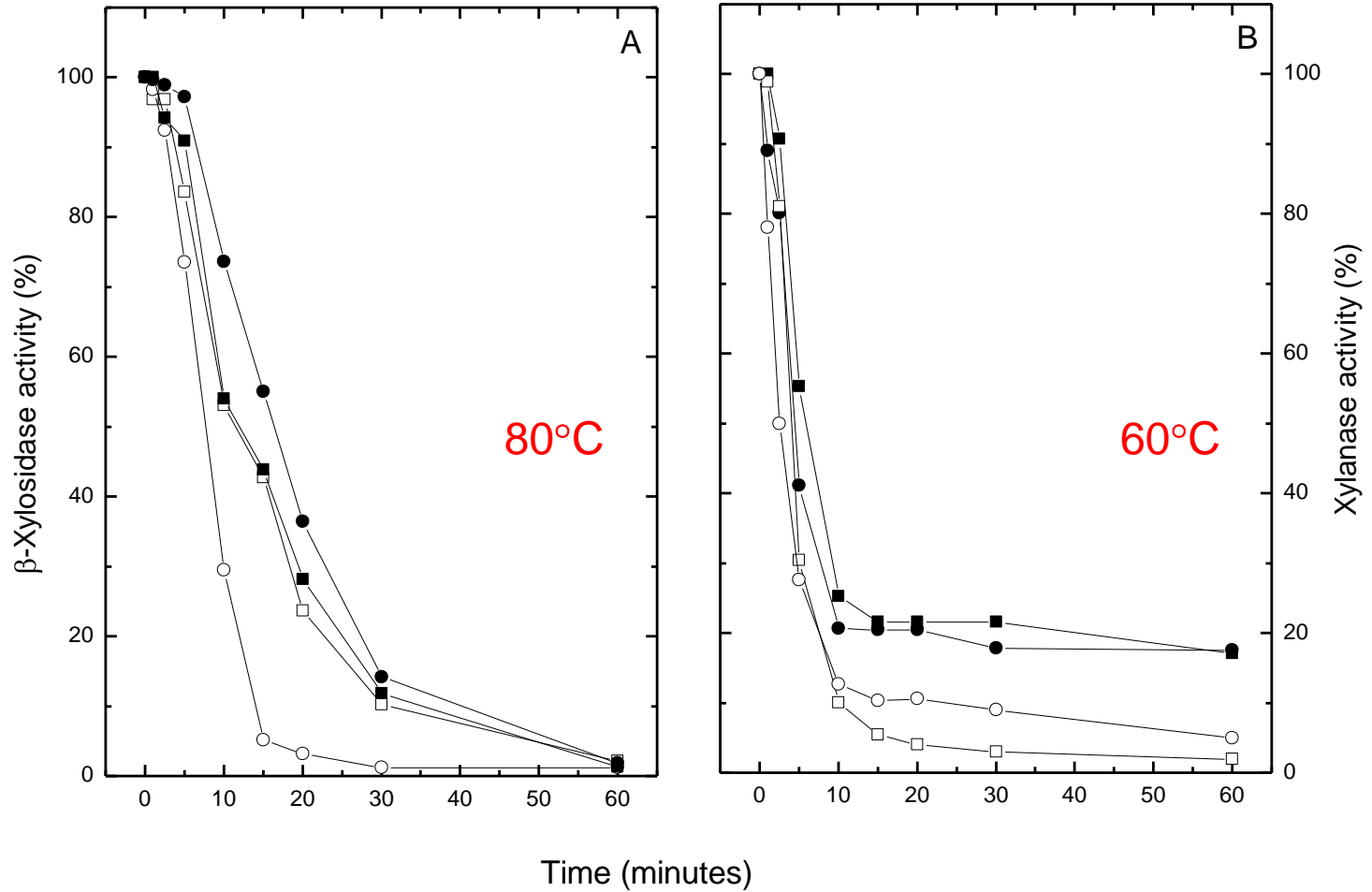
extracellular

intracellular



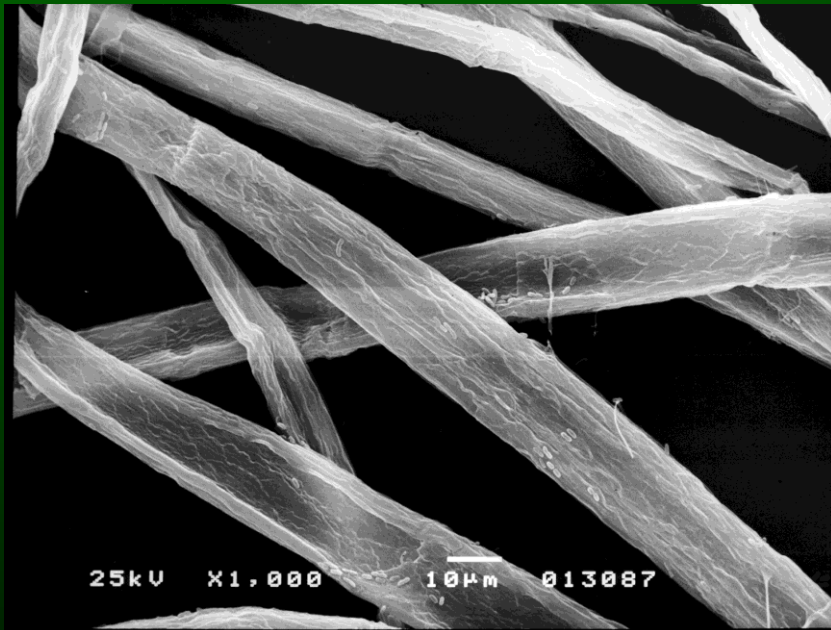
25°C ( $\square$ ,  $\circ$ ), 42°C ( $\blacksquare$ ,  $\bullet$ )

# Thermal inactivation

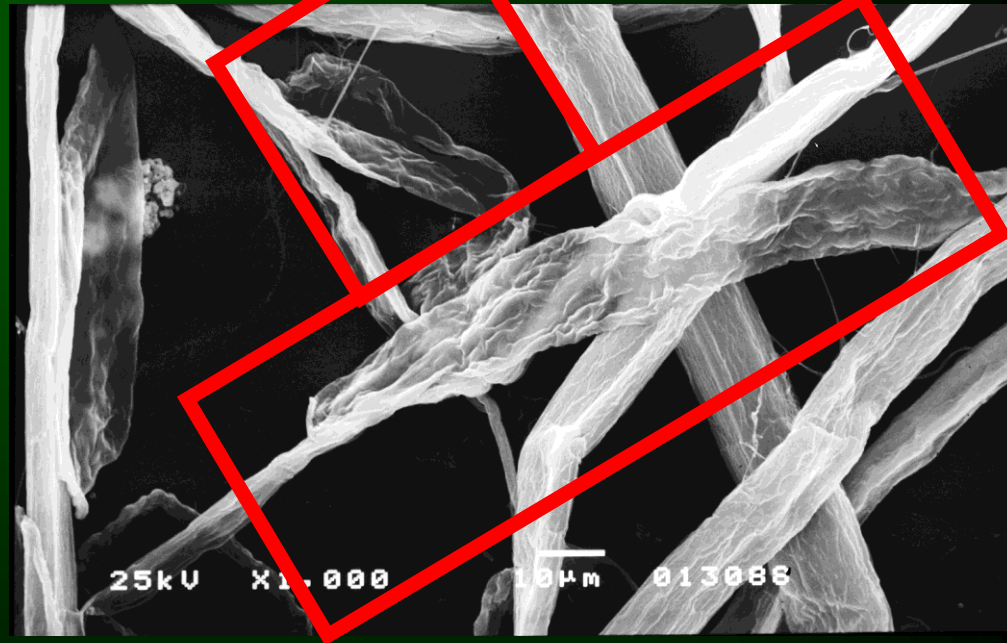


25°C ( $\square$ ,  $\circ$ ); 42°C ( $\blacksquare$ ,  $\bullet$ ); extra ( $\square$ ), intra ( $\circ$ )

# ELECTRONIC MICROSCOPY OF CELLULOSE TREATED WITH XYLANASE

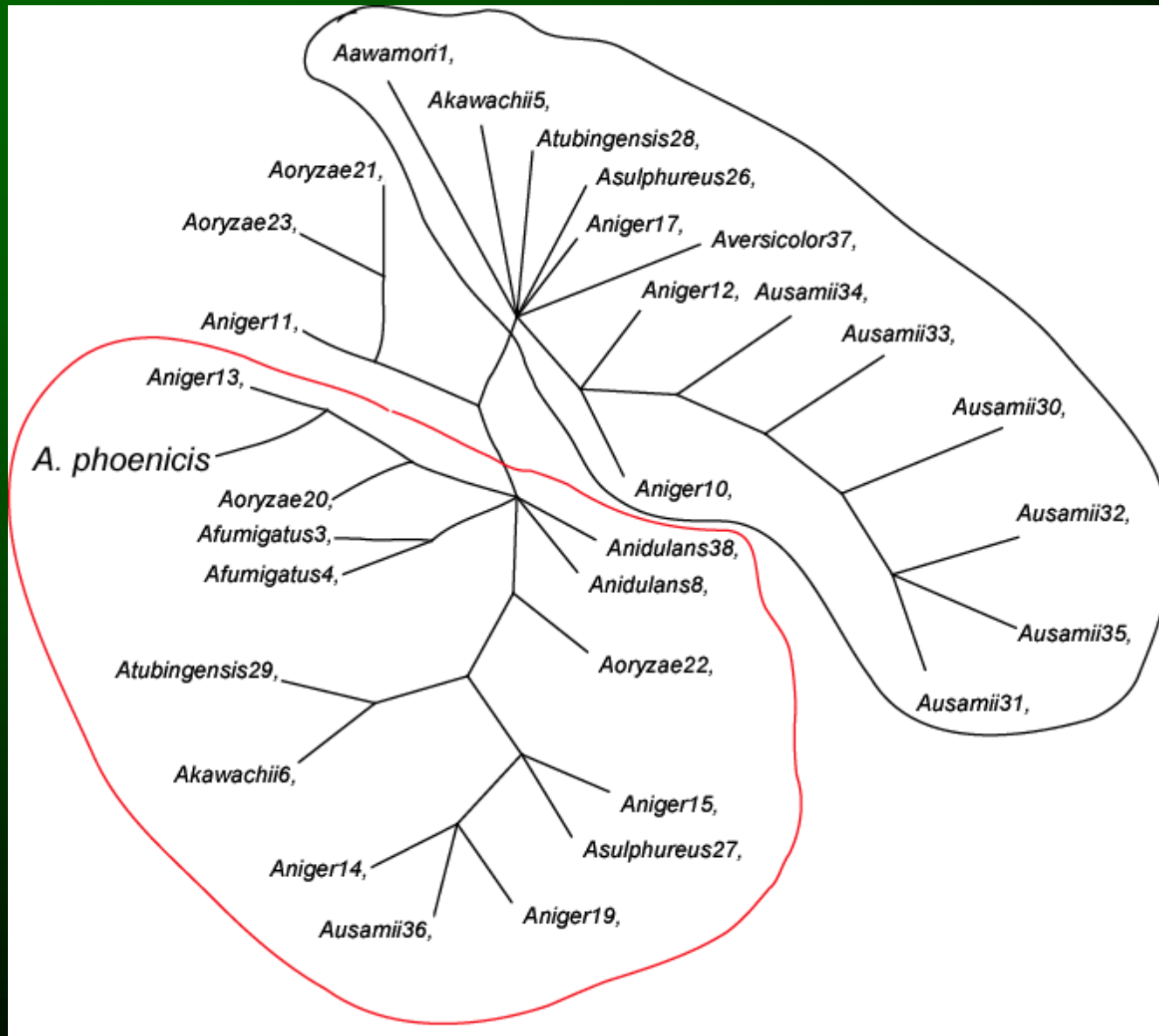


control



2 hours of incubation Xylanase  
15 U/g dry cellulose pulp

# PHYLOGENETIC RELATIONSHIPS AMONG GENUS *Aspergillus*



Corrêa, E. C. et al. (submitted to Molecular Biology and evolution) 2008.



# Perspectives

## Development of Enzymes for Biomass Saccharification

Overexpression of  
enzymes in heterologous  
organisms

Production of enzymes  
and fungi quick adaptation  
(directed anagenetic  
evolution)

Comparative and  
evolutionary approaches of  
different groups of enzymes

purification and  
biochemical  
characterization of  
microbial enzymes and  
sequencing

**OUR**

**LABORATORY**

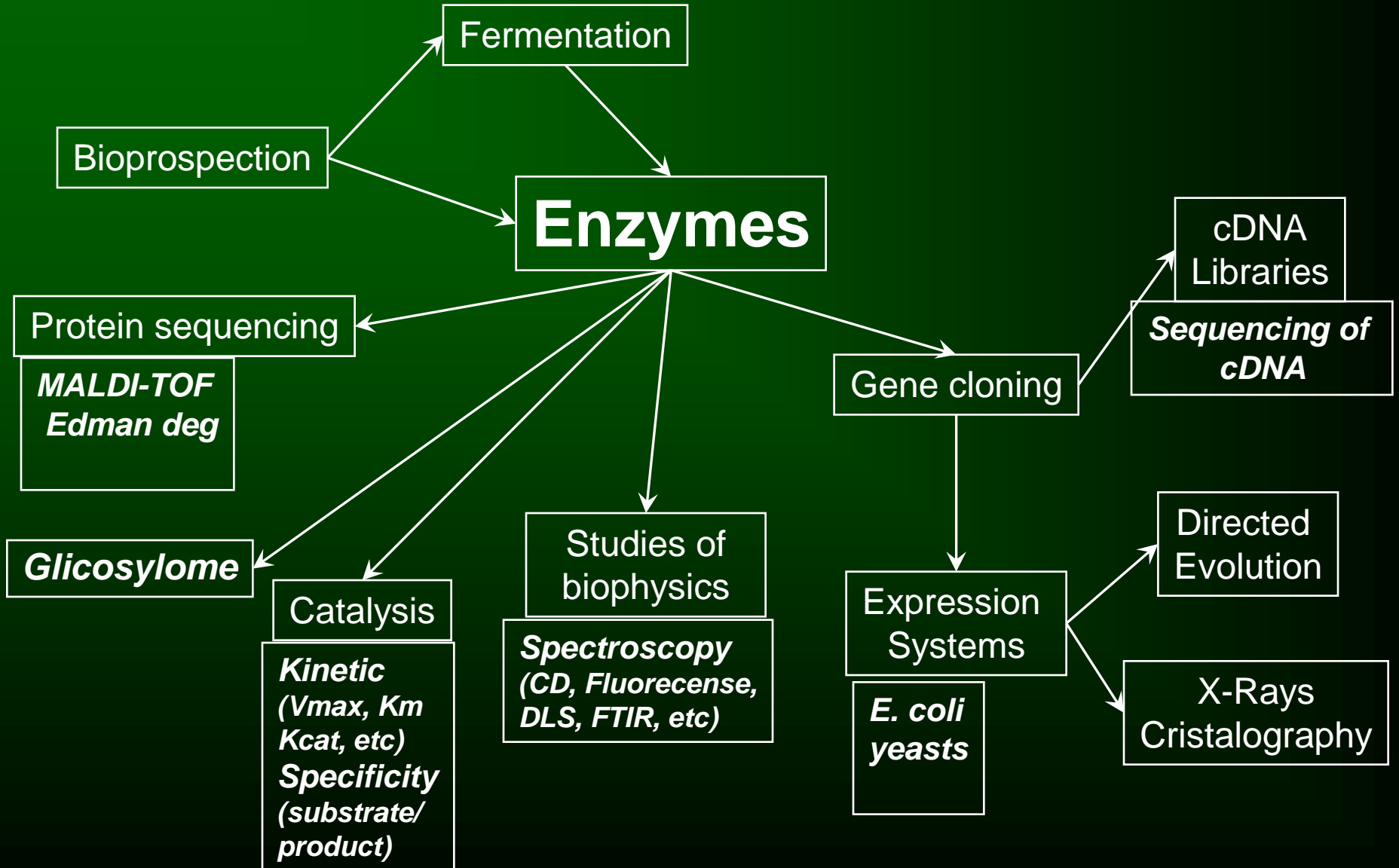
Bioprospection of  
filamentous fungi

Sparse matrix techniques  
for optimization of  
microbial enzyme  
production

Submerged and solid-  
substrate fermentations  
(using residues from agro-  
farming and forestry  
industries)

catabolic induction and  
repression studies using  
diverse carbon sources

# PLAN



# Research Group





*Thank you*

